

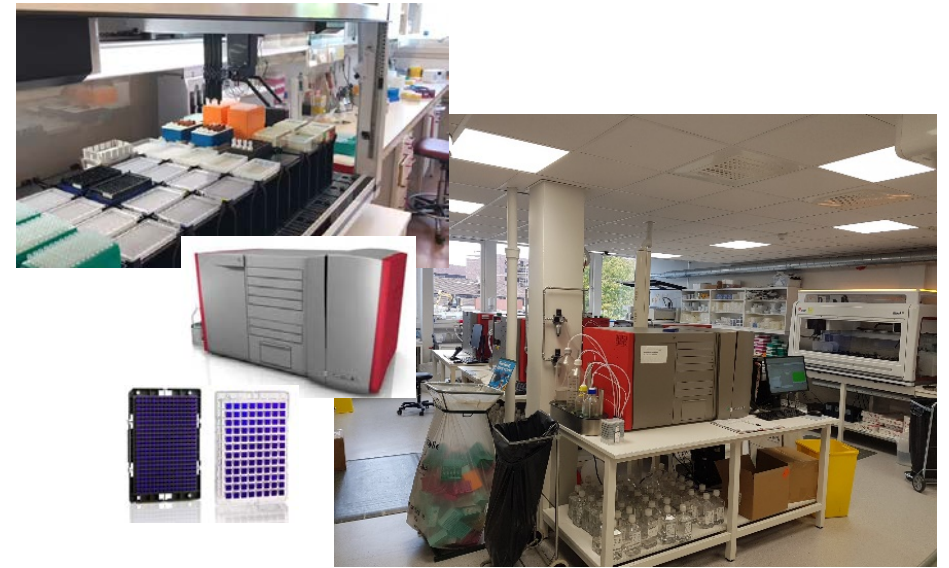
CIGENE lab



- ✓ Established 2003 as a FUGE platform
- ✓ High throughput SNP-genotyping facility

Well equipped genomics lab:

- ✓ Sequencing
- ✓ Functional genomics assays
- ✓ Gene editing (CRISPR)
- ✓ Cell lab



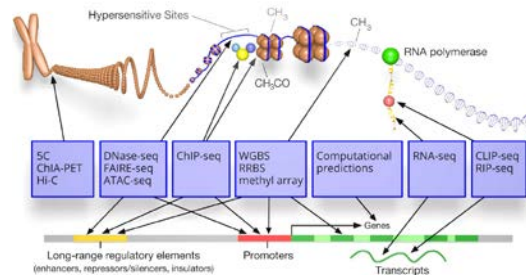
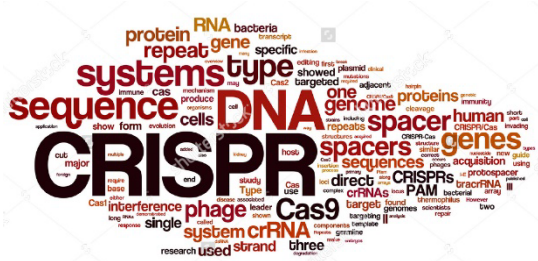
**Genome biology,
IHA, BIOVIT**

Muliggjørende verktøy og teknologier

'Long-read' sekvensering



Funksjonell annotering for å forstå genomfunksjon



DIGITAL SALMON





- Lønn til labpersonell og andre personer tilknyttet labdrift
- Oppgradering av utstyr:
 - 2017 -> 8,1 mill.
 - 2018 -> 3,8 mill.
 - 2019 -> 1,1 mill.
- Servicekontrakter (3 mill. pr. år)
- Investering i nye teknologier og metoder



- Genomsekvensering
- Funksjonell annotering
- Bioinformatikk
- Geneditering (CRISPR)

CIGENE lab

ORION Computer cluster



**Faggruppe genombiologi,
IHA, BIOVIT**



2018

- Oppgradering av CIGENE lab
- 2 mill. kr

Budsjett 2019

Genotyping	Prøver	Omsetning	Consumables	Inntekt
Aviagen, AquaGen	450 000	22 500 000	6 300 000	16 200 000
Agena Genotyping	6 500	236 979	148 958	88 021
SUM			6 448 958	16 288 021
Ansatte	Stillinger	Lønn	Andel	Lønnsutgifter
CIGENE, labstab	7,85	692 114		-5 433 097

Oppdragsaktivitet

Forskere

Personale

Sum lønn				-7 469 476
Servicekontrakt				-3 000 000
Diverse utviklingskostn. lab.				-1 000 000
Lab fee				-300 000
Kompetanseheving CIGENE labstab				-100 000
Nytt utstyr				
Beckman i5 S8 pipetteringsrobot				-875 000
Binder kjøleinkubator				-70 000
Mikrosentrifuger og vortex ny lab				-35 000
Pipetter til ny lab				-55 000
Stoler til ny lab				-80 000
Sum utstyr				-1 115 000
Lønn fra REWIRED				214 575
FILOPAT	2 000	250 000	28 000	222 000
QE2	3 000	375 000	42 000	333 000
EcoEvoGene	2 000	250 000	28 000	222 000
Primmer	1 500	187 500	21 000	166 500
Nanopore sequencing				60 000
Markørpanel for Arktisk røye				400 000
Andre prosjekt inntekter				500 000
SUM				2 118 075
TOTAL				5 421 620

Driftskostnader

Nytt utstyr

NFR-prosjekter

Prof., ass.prof., prof. II



Sigbjørn Lien



Dag Inge Våge



Simen Sandve



Guro Sandvik



Jon Olav Vik
25%



Øivind Andersen
20%



Lise Fjellsbø

Adm.



Researchers



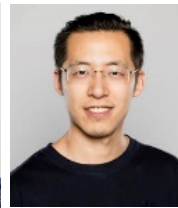
Matthew
Kent



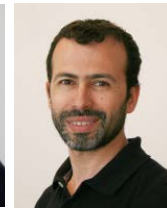
Nicola
Barson



Torfinn
Nome



Yang Jin



Amine
Namouchi



Hien To



Sahar
Hassani



Marie
Gulla



Filip
Rotnes



Gareth
Gillard



Jørn Henrik
Sønstebo



Victor
Boyartchuk



Fabian 33%
Grammes

Postdocs



Lars
Grønvold



Marie-Odile
Baudement



Michel
Moser



Thomas
Harvey



Olga
Paulouskaya



Tomasz
Podgorniak



Tina G.
Kirubakaran



Mathilde
Holen



Line
Røsæg



Øystein
Monsen



Live Rud-
Johansen



Darshan
Young



Kristina
Stenløkk



Noman
Reza

PhD-students

Technicians



Kristil
Sundaasen



Silje
Karoliussen



Mariann
Arnyasi



Therese
Andersstuen



Tan Thi
Nguyen



Øystein
Milvang



Kristin Udjus



Nina Falk
Peterson

+

- 3 technicians
- 2 Phd
- 5 postdocs

> 50 people

Prof., ass.prof., prof. II



Sigbjørn Lien



Dag Inge Våge



Simen Sandve



Guro Sandvik



Jon Olav Vik
25%



Øivind Andersen
20%



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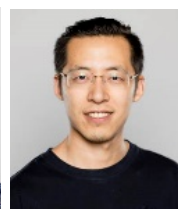
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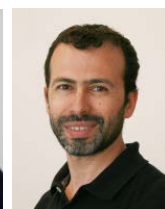
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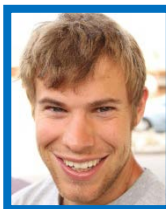
Postdocs



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- 5 postdocs

> 50 people

Ny master i «Genome Science» (genomvitenskap)

- ❑ Rekruttere studenter fra bachelorstudiene i biologi, bioteknologi, plante-, husdyr- og 'havbruk'/akvakultur, samt studenter fra andre læresteder
- ❑ Opptak med krav om «grunnpakke» av relevante kurs innen genetikk/genomikk, matematikk/statistikk/informatikk, biokjemi/molekylærbiologi/bioteknologi...
- ❑ Bred innretning mot bioinformatikk, funksjonell genomikk, genombasert avl, epigenetikk, genomevolusjon, ernæringsgenomikk, mikrobiota, mikroorganismer, patogener, ...
- ❑ **BIOVIT:** Genombiologi, Avl og kvantitativ genetikk, Genetikk og planteforedling, Plantebiologi og bioteknologi. **KBM:** Biostatistikk, bioinformatikk, mikrobiota/mikrobiell genomikk. **REALTEK:** Data Science. **VET:** Sykdomsgenetikk, patogener, diagnostikk?
- ❑ Introduksjonskurs som kombinerer data-generering (lab) med data-analyse
- ❑ **Labkurs i funksjonell genomikk/genredigering/sekvensering**
- ❑ Nye kurs med appell for studenter også på andre studieretninger

World leading on salmonid genomics

ARTICLE

OPEN

doi:10.1038/nature17164

The Atlantic salmon genome provides insights into rediploidization

Sigbjørn Lien¹, Ben F. Koop², Simen R. Sandve¹, Jason R. Miller³, Matthew P. Kent¹, Torfinn Nome¹, Torgeir R. Hvidsten^{4,5}, Jong S. Leong², David R. Minkley², Aleksey Zimin⁶, Fabian Grammes¹, Harald Grove¹, Arne Gjuvsland¹, Brian Walenz³, Russell A. Hermansen^{7,8,9}, Kris von Schalburg², Eric B. Rondeau³, Alex Di Genova^{10,11}, Jeevan K. A. Samy¹, Jon Olav Vik¹, Magnus D. Vigeland¹², Lis Caler¹, Unni Grimholt¹³, Sissel Jentoft¹⁴, Dag Inge Våge¹, Pieter de Jong¹⁵, Thomas Moen¹⁶, Matthew Baranski¹⁷, Yniv Palti¹⁸, Douglas R. Smith^{19,20}, James A. Yorke⁶, Alexander J. Nederbragt¹⁴, Ave Tooming-Klunderud¹⁴, Kjetill S. Jakobsen¹⁴, Xuanting Jiang²¹, Dingding Fan²¹, Yan Hu²¹, David A. Liberles^{8,9}, Rodrigo Vidal²², Patricia Iturra²³, Steven J. M. Jones^{24,25}, Inge Jonassen²⁶, Alejandro Maass^{10,11}, Stig W. Omholt²⁷ & William S. Davidson²⁵

The whole-genome duplication 80 million years ago of the common ancestor of salmonids (salmonid-specific fourth vertebrate whole-genome duplication, Ss4R) provides unique opportunities to learn about the evolutionary fate of a duplicated vertebrate genome in 70 extant lineages. Here we present a high-quality genome assembly for Atlantic salmon (*Salmo salar*), and show that large genomic reorganizations, coinciding with bursts of transposon-mediated repeat expansions, were crucial for the post-Ss4R rediploidization process. Comparisons of duplicate gene expression patterns across a wide range of tissues with orthologous genes from a pre-Ss4R outgroup unexpectedly demonstrate far more instances of neofunctionalization than subfunctionalization. Surprisingly, we find that genes that were retained as duplicates after the teleost-specific whole-genome duplication 320 million years ago were not more likely to be retained after the Ss4R, and that the duplicate retention was not influenced to a great extent by the nature of the predicted protein interactions of the gene products. Finally, we demonstrate that the Atlantic salmon assembly can serve as a reference sequence for the study of other salmonids for a range of purposes.

200 | NATURE | VOL 533 | 12 MAY 2016

LETTER

doi:10.1038/nature16062

Sex-dependent dominance at a single locus maintains variation in age at maturity in salmon

Nicola J. Barson^{1,6}, Tutku Aykanat^{2,6}, Kjetil Hindar³, Matthew Baranski⁴, Geir H. Bolstad³, Peder Fiske³, Céleste Jacq⁴, Arne J. Jensen³, Susan E. Johnston⁵, Sten Karlsson¹, Matthew Kent¹, Thomas Moen⁶, Eero Niemelä⁷, Torfinn Nome¹, Tor F. Næsje³, Panu Orell¹, Atso Romakkaniemi⁷, Harald Sægvog⁸, Kurt Urdal⁸, Jaakko Erkinaro⁷, Sigbjørn Lien¹ & Craig R. Primmer²



17 DECEMBER 2015 | VOL 528 | NATURE | 405

New Results

Comment on this paper

Title: Sex-dependent dominance maintains migration supergene in rainbow trout

Devon E. Pearse, Nicola J. Barson, Torfinn Nome, Guangtu Gao, Matthew A. Campbell, Alicia Abadía-Cardoso, Eric C. Anderson, David E. Rundio, Thomas H. Williams, Kerry A. Naish, Thomas Moen, Sixin Liu, Matthew Kent, David R. Minkley, Eric B. Rondeau, Marine S. O. Briec, Simen Rød Sandve, Michael R. Miller, Lucydalla Cedillo, Kobi Baruch, Alvaro G. Hernandez, Gil Ben-Zvi, Doron Shem-Tov, Omer Barad, Kirill Kuzishchin, John Carlos Garza, Steven T. Lindley, Ben F. Koop, Gary H. Thorgaard, Yniv Palti, Sigbjørn Lien

doi: <https://doi.org/10.1101/504621>

This article is a preprint and has not been peer-reviewed [what does this mean?]

Abstract

Full Text

Info/History

Metrics

Preview PDF

Abstract

Traits with different fitness optima in males and females cause sexual conflict when they have a shared genetic basis. Heteromorphic sex chromosomes can resolve this conflict and protect sexually antagonistic polymorphisms but accumulate deleterious mutations. However, many taxa lack differentiated sex chromosomes, and how sexual conflict is resolved in these species is largely unknown. Here we present a chromosome-anchored genome assembly for rainbow trout (*Oncorhynchus mykiss*) and characterize a 56 Mb double-inversion supergene that mediates sex-specific migration through sex-dependent dominance, a mechanism that reduces sexual conflict. The double-inversion contains key photosensory, circadian rhythm, adiposity, and sexual differentiation genes and displays frequency clines associated with latitude and temperature, revealing environmental dependence. Our results constitute the first example of sex-dependent dominance across a large autosomal supergene, a novel mechanism for sexual conflict resolution capable of protecting polygenic sexually antagonistic variation while avoiding the homozygous lethality and deleterious mutation load of heteromorphic sex chromosomes.

Nature Ecology & Evolution, (in press)